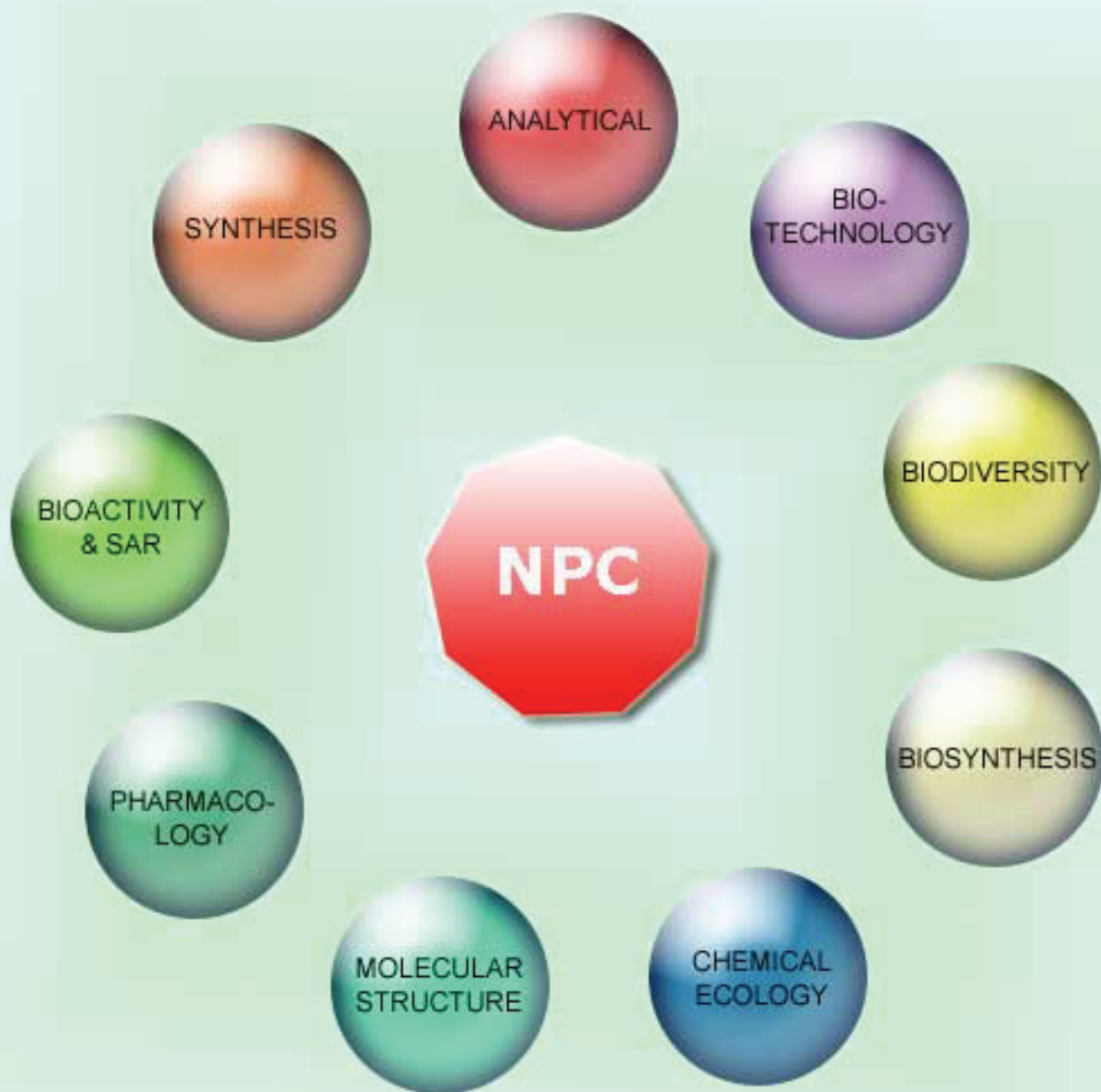


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Pteridaceae Fragrant Resource and Bioactive Potential: a Mini-review of Aroma Compounds

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Seven ferns of *Pteridaceae*, grown in a botanical garden or wild, harvested in France were investigated for their Volatile Organic Compounds (VOC) profile using GC-MS: *Adiantum pedatum* L., *Adiantum peruvianum* Klotzsch, *Anogramma leptophylla* (L.) Link, *Cheilanthes maderensis* Lowe, *Cryptogramma crista* (L.) R. Br., *Pteris cretica* L. and *Pteris vittata* L. Fifty-three VOC biosynthesized from lipidic, shikimic, terpenic and carotenoid pathways were identified. The two *Adiantum* species show different VOC composition. The main linalool (10.8%) in *A. pedatum* has several biological activities of great interest. This Maidenhair fern contains the highest proportion (57.9%) of isoprenoid flavor precursors, i.e., ionone derivatives with various scent notes. The two major odorant unsaturated hexenoic acids derivatives of *A. peruvianum* are used as flavouring agents. *Anogramma leptophylla* concentrates 6-methoxymellein (71.5%), a bitter phytoalexin which contributes to stress or pathogen resistance. *Cheilanthes maderensis* produces mainly coumarin (89%) and vanillin (3.5%) with a low odor detection threshold, both used in perfumery and cosmetic industry or as flavouring agent and drug additives. *Cryptogramma crista* accumulates a broad-spectrum of carotenoid derivatives (52.1%) and three major shikimic derivatives: the spicy 4-vinylguaiaicol (flavouring agent), the floral phenylethanal and benzyl alcohol with floral, balsamic scent. *Pteris cretica* accumulates mostly furan derivatives, i.e., 5-hydroxymethylfurfural (33.2%) and 3-hydroxy-2,3-dihydromaltol (18.3%) used as food and beverage additives with caramel or roasty flavour and also found in fortified wines, toasty or heat-treated foods. *Pteris vittata* produces predominantly shikimic derivatives applied in perfumery and food industries as benzaldehyde (26%, with almond scent), benzyl alcohol (22%, floral fruity balsamic scent), nonanal (19.8% cucumber note) and phenylethanal (11%; floral note). *Pteridaceae* resources are of great interest as a reservoir of odorous and bioactive compounds.

Keywords: Benzaldehyde, Coumarin derivatives, Furan derivatives, Linalool, 6-Methoxymellein, Nonanal, 4-Vinylguaiaicol.

Pteridaceae E. D. M. Kirchner is a heterogeneous family of ferns including approximately nine hundred species worldwide distributed. Only twelve species belonging to seven genera (*Adiantum* L., *Anogramma* Link, *Cheilanthes* Swartz, *Cosentinia* Tod., *Cryptogramma* R. Br. ex Richardson, *Notholaena* R. Br. = *Paragymnopteris* K.H. Shing and *Pteris* L.) are reported in France [1a,b].

Pteridaceae includes several species well-known in traditional medicines and non-pharmacological interventions, i.e., Native American people medicine (Navajo Indian Tribe), Ayurvedic medicine, homeopathy linked with various diseases or disorders as follows. Leaves of *Adiantum pedatum* L. are employed for pectoral affections, *Adiantum poiretii* Wickstr. against fever and diabetes, *Notholaena eckloniana* Kunze (L.) as ointment on the scalp and *Pteris wallichiana* J. Agardh applied to stop bleeding. Fronds of *Pellaea calomelanos* (Sw.) Link are used against asthma; those of *Pteris multifida* Poir. and *Pteris cretica* L. are applied against dysentery and wounds, respectively [1c-f]. Leaves of *Adiantum capillus-veneris* L. (also known as Venus-hair fern) are used against throat affections and, as purgative and demulcent. This fern is also the main ingredient of the renowned "Sirop de Capillaire" supposed to cure a large number of diseases [1f]. On the other hand, Venus-hair fern produces a pleasant tonic flavor and syrups which are used as a flavor modifier [1f]. Rhizomes of *Cheilanthes tenuifolia*

(Burm. f.) Sw., are administered as general tonic while those of *Pteris ensiformis* Burm. f. and *Pteris quadriaurita* Retz. are applied on swollen glands in the neck or healing of boils, respectively. It should be mentioned *Adiantum lunulatum* Burm. to be used against fever due to elephantiasis. Finally root of *Cheilanthes farinosa* (Forsk.) Kaulf. treats eczema and stomachache [1c-f].

During the last decades several studies have been carried out regarding the biological properties of *Pteridaceae*: an antioxidant activity was detected in *Adiantum trapezifolium* L. and *C. tenuifolia* [2a,b] and antimicrobial compounds were characterized in *Pteris vittata* L. and *Pteris biaurita* L. [2c,d]. Biological activities involved in metabolic syndrome and anti-tumor activity were investigated for *P. vittata* [2e,f]. The impact of heavy metals on antioxidant polyphenols of this hyperaccumulator fern was also analysed [2g]. In addition, an anti-tumor activity was detected in *Pteris semipinnata* L. and *Pteris multifida* [2h,i] while an aqueous extract of *Pteris ensiformis* demonstrates an immunomodulatory activity [2j].

Very few *Pteridaceae* are known having an odor [3a]: only *Adiantum pantadactylon* Langsd. & Fisch. and *Pteris tremula* R. Br. are reported to smell tom cat urine. *Pteris multifida* has an acrid and biting flavor whereas *Adiantum pedatum* has a slightly aromatic odor.

Table 1: Percentage of volatile organic compounds^a in fresh aerial part of seven species of *Pteridaceae*.

Compounds	RI ^b	<i>Adiantum pedatum</i>	<i>Adiantum peruvianum</i>	<i>Anogramma leptophylla</i>	<i>Cheilanthes maderensis</i>	<i>Cryptogramma crispa</i>	<i>Pteris cretica</i>	<i>Pteris vittata</i>
Lipidic derivatives								
Furfural	840						3.0	
Furfuryl alcohol	910						1.2	
5-Methylfurfural	965						2.1	
1-Octen-3-ol	976	5.0	7.1			5.6	3.9	9.5
2,3-Octanedione	979		2.0					
3-Octanol	990		1.6				1.1	
(<i>E</i>)-3-Hexenoic acid	1043		23.3					
(<i>E</i>)-Oct-2-en-1-ol	1067		0.8					
(<i>E</i>)-2-Hexenoic acid	1068		19.0					
Furanol	1083						3.8	
Nonanal	1107	0.8	0.8	0.8	1.0	0.7	1.6	19.8
5-Hydroxymethylfurfural	1273						33.2	
6-Methoxymellein	1893			71.5				
NI	1907			1.7				
6-Hydroxymellein	1947			1.8				
NI ^c	1980			1.0				
Shikimic derivatives								
Benzaldehyde	962	2.4		7.7		3.3	1.2	26.0
Benzyl alcohol	1034	5.6	4.6			6.1	1.2	22.0
Phenylethanal	1045	1.9		1.0		8.8	0.8	11.0
Acetophenone	1065							4.0
2-Phenylethanol	1118		2.9			3.2		
3,5-Dihydroxy-6-methyl-2,3-dihydro-4 <i>H</i> -pyran-4-one	1169						18.3	
4-Ethenyl-2-methoxyphenol	1316		0.6			10.0	2.7	
NI	1320	2.4						
2,3-Dihydrocoumarin	1389		1.1	0.3	3.4			
4-Hydroxy-3-methoxybenzaldehyde (vanillin)	1402	1.6	1.7		3.5	3.5	1.9	6.1
Coumarin	1443		1.2	7.8	89.0			
4-Hydroxybenzoic acid	1575					3.2		
4-Hydroxy-3-methoxybenzoic acid (vanillic acid)	1603					2.1		0.3
4-Hydroxy-3-methoxyacetophenone	1743			3.5				
6,7-Dimethoxycoumarin (scoparone)	1775			0.8				
7-Methoxy-6-prenylcoumarin (suberosin)	1970			1.0				
Terpenic derivatives								
Linalool	1102	10.8	0.6					
(<i>Z</i>)-2,6-Dihydroxy-2,6-dimethyl-3,7-octadiene	1194	4.6	1.4					
2,6-Dimethyl-3,5,7-octatriene-2-ol (<i>E,E</i> or <i>E,Z</i>)	1212	3.9						
8-Hydroxylinalool	1369	2.6	1.4					
Carotenoid derivatives								
3,4-Dehydro-7,8-dihydro- β -ionone	1413	2.4				1.6	0.4	
α -Ionone	1428		0.6					
NI	1434					1.2	2.4	
NI	1457						0.9	
3,4-Dehydro- β -ionone	1475						1.1	
β -Ionone	1477		1.2				0.5	
NI	1486						0.7	
10-Methyl- α -ionone	1492						0.7	
NI	1520					4.8		
Dihydroactinidiolide	1527		0.4			0.2		
8-Methyl- α -ionone	1548					8.8		
NI	1580	0.4						
(<i>E,E</i>)-Pseudoionone	1584	11.0					0.5	
NI	1605		6.1				0.5	
NI	1615	3.0						
3-Hydroxy- β -damascone	1616		1.2				0.5	
3-Hydroxy- β -ionol	1630	3.0						
3-Hydroxy-7,8-dehydro- α -ionol	1633		1.2				0.4	
9-Methyl- α -ionol	1637	5.0	0.6			9.2	2.5	0.2
3-Oxo- α -ionol	1650	4.0	1.2		0.3	5.2	1.5	
4-Hydroxy-7,8-dihydro- β -ionone	1651	3.6	3.1			6.8	1.5	
3-Hydroxy- β -ionone	1664	3.6					2.6	
4-Hydroxy-5,6-epoxy-ionol	1675	13.4	3.2		1.6	1.1	4.5	
3-Hydroxy-7,8-dihydro- β -ionol	1678	2.0	2.6					
3-Hydroxy-5,6-epoxy-ionone	1691	6.5	4.4			12.2	1.6	0.3
2-Hydroxy- β -ionone	1693		1.9			1.0		
3-Oxo-6-hydroxy- α -ionone	1796		0.6					

^a Relative percentage of the VOC based on the GC-MS chromatographic area; ^b RI = Retention Indices on SLBTM-5MS column (Supelco); ^c NI = Non Identified

With a view to continue our study of Volatile Organic Compounds (VOC) with bioactive potential, fresh aerial parts of seven ferns of *Pteridaceae* harvested in France were investigated for their VOC profile using GC-MS: *Adiantum pedatum* L., *Adiantum peruvianum* Klotzsch, *Anogramma leptophylla* (L.) Link, *Cheilanthes maderensis* Lowe, *Cryptogramma crispa* (L.) R. Br., *Pteris cretica* L. and *Pteris vittata* L.

In the concentrated diethyl ether extracts of the seven species, fifty-three components biosynthesized from lipidic, shikimic, terpenic and carotenoid pathways were identified (Table 1). The volatile fraction of the ferns represents about 0.01% of the fresh aerial materials.

Twenty volatile compounds were identified in *Adiantum pedatum*. Lipidic derivatives are mainly represented by 1-octen-3-ol (5%) responsible for the mushroom-like odor and flavor [3b-d] but also found in many plants [3e-g]. This fatty alcohol is valuable to perfume and food industries [3h, 4a,b] and more recently proposed for mosquito control as an insect attractant [4c,d]. Benzyl alcohol (5.6%), the major compound of the shikimic pathway with floral odor also described as phenolic or balsamic [4a] and the main terpenic derivative linalool (10.8%) with floral, fruity scent [4c] or woody note (depending on the enantiomer) were also reported in table 1. Linalool is a well-known terpenic alcohol of essential oil from various plant families (*Lauraceae*, *Rutaceae*, *Lamiaceae*...); it gives insect repellent property as well as anxiolytic, anti-

inflammatory, antioxidant, antifungal, antibacterial, antiparasitic, antitumoral activities [4e-i]. This fern contained the highest proportion of isoprenoid flavor precursors (57.9%), i.e., mainly (*E,E*)-pseudoionone (11%) with odor descriptors as sweet, waxy, citrus, floral balsamic, spicy [4a,j], 4-hydroxy-5,6-epoxy-ionol (13.4%), 3-hydroxy-5,6-epoxy- β -ionone (6.5%) and 9-methyl- α -ionol (5%).

Adiantum peruvianum showed a VOC profile based on twenty-nine compounds, radically different from the previous *Adiantum* species: lipidic derivatives (54.6%) were mainly represented by (*E*)-3-hexenoic acid (23.3%) with honey odor and waxy, fruity or herbal notes [4a], (*E*)-2-hexenoic acid (19%) with fruity odor and 1-octen-3-ol (7.1%) with mushroom-like scent. The two major odorant hexenoic acids, used as flavouring agents, were previously found in other ferns such as *Athyrium filix-femina*, *Gymnocarpium dryopteris*, *Polystichum setiferum*, *Pteridium aquilinum* [5a,b] and plant allies (*Equisetum palustre*) [5c] but not in *Adiantum capillus-veneris*. In Venus-hair fern, (*E*)-2-decenal, lauric amide or (*E*)-2-heptenal were found in high quantities with a plastic or oxidized mutton fat odor [5a], also responsible for the unpleasant scent of “stink bug”. The VOC profiles of the three species of *Adiantum* are therefore different.

Carotenoid derivatives of *A. peruvianum* (28.3%) were composed by small amounts of α -ionone, β -ionone and ionone derivatives, i.e., 3-hydroxy-5,6-epoxy- β -ionone. The VOCs from the shikimic pathway (12.1%) were represented by few compounds including benzyl alcohol (4.6%) also described in *A. pedatum* (Table 1) as well as 2-phenylethanol, vanillin or coumarin previously found in *A. pedatum*, *A. peruvianum* and *A. trapeziforme* [5d]. Three minor terpenic compounds including linalool were also identified in *A. peruvianum* (3.4%). This second *Adiantum* species, as well as the previously analysed Venus-hair fern, produced small amounts of terpenic derivatives. The five other ferns analysed in this work and belonging to four other genera of *Pteridaceae* did not produce any terpenic derivatives.

Ten VOCs were detected from *Anogramma leptophylla*. The volatile pattern was mainly based on lipidic derivatives (76%), i.e., the major 6-methoxymellein (71.5%) and the minor 6-hydroxymellein (1.8%) which are 3,4-dihydroisocoumarins. The former is a polyketide-derived phytoalexin well-known in the carrot and would contribute to pathogen or stress resistance. It is the first compound related to the bitterness of the carrot and its content varies in the commercial products with storage and processing conditions [6a-c]. Dihydroisocoumarins have been isolated from other plants species and also from macrofungi [6d,e]. The others VOC isolated from *A. leptophylla* were shikimic derivatives (22.1%), i.e., benzaldehyde (7.7%) widespread in plants and mushrooms with bitter almond odor and coumarin (7.8%) with pleasant scent. These VOCs are two aroma agents commonly used in perfume, cosmetic and food industries.

The volatile content of *Cheilanthes maderensis* was mainly dominated by shikimic derivatives (95.9%) essentially coumarin (89%: hay and dried herb odor), 3,4-dihydrocoumarin (3.4%; hay-like, herbal, coconut note) and vanillin (3.5%; vanilla, sweetish smell) usually used in perfume and food industries [3h; 4a,b]. Such high content of coumarin and coexistence of its dihydro derivative as natural products are very rare. Recently, a Japanese group reported the similar data from the bryophyte, *Takakia lepidozoioides* [3i], as those reported in the present paper. At the same time, chemophylogenetic relationship between both phyla (Pteridophytes and Bryophytes) has been fully discussed [3j].

The broad spectrum of volatile components identified in *Cryptogramma crispera* showed a VOC profile including nineteen identified compounds. Table 1 lists major carotenoid derivatives (52.1%), i.e., 3-hydroxy-5,6-epoxy-ionone, 9-methyl- α -ionol, 8-methyl- α -ionone, 4-hydroxy-7,8-dihydro- β -ionone and 3-oxo- α -ionol. Shikimic derivatives (40.2%) are mainly represented by three VOCs. 4-Ethenyl-2-methoxyphenol also called 2-methoxy-4-vinylphenol or 4-vinylguaiaicol (10%) with powerful, clove-like, spicy, smoky odor is also a flavouring agent and a pheromone for insects [4b,j; 7a,b]. It was previously found in a horsetail, *Equisetum telmateia* [5c]. Phenylethanal (8.8%) with floral odor (lilac, hyacinth, geranium: [3b, 4b]) was also identified in other ferns and plant allies (*Athyrium filix-femina*, *Blechnum spicant*, *Phegopteris connectilis*, *Equisetum scirpioides*) [5a-c]. The third shikimic derivative was benzyl alcohol (6.1%) with floral or balsamic odor. Only 1-octen-3-ol from lipidic pathway was identified in a significant amount (Table 1).

Table 1 lists a broad spectrum of VOCs for *Pteris cretica* based on twenty-seven identified components mainly lipidic derivatives (49.9%), shikimic derivatives (26.1%) and carotenoid derivatives (22.8%) in low amounts. Only two VOCs were abundant as 5-hydroxymethylfurfural (33.2%) and 3,5-dihydroxy-6-methyl-2,3-dihydro-4*H*-pyran-4-one (18.3%). The major 5-hydroxymethylfurfural is described with odor of chamomile flowers (or butter, caramel, musty) while the four others furan derivatives (10%) have various descriptors [8a-d] as furfural (almond, woody, sweet, toasty), furfuryl alcohol (faint burning odor), furaneol (sweet, caramel, pineapple, strawberry) and 5-methylfurfural (caramel, almond, spicy, sweet, roasty). These compounds usually found in fortified wines, in roasted, toasted or heat-treated foods and drinks, are produced, in particular, by sugar alteration (Maillard reaction). Suspected but not proved to be carcinogenic, they contribute to caramel aroma and colour in food additives [8e-h]. 3,5-Dihydroxy-6-methyl-2,3-dihydro-4*H*-pyran-4-one (or 3-hydroxy-2,3-dihydromaltol 18.3%, Table 1), the second major VOC exhales an odor with toasty character and fruity-caramel overtones [7a].

Pteris cretica was the single species of the seven analysed ferns with a high level of furaneol and furfural derivatives (43.2%, Table 1) whereas the others (in particular *P. vittata*) contain none at all. Furan derivatives were not found in *A. capillus-veneris*, another species from *Pteridaceae* previously studied [5a]. However, ferns species from other families may also produce furan derivatives in small amounts, i.e., *Pteridium aquilinum*, *Asplenium trichomanes*, and the twelve species of the *Asplenoideae* family [5b,9a]. Other authors found furan derivatives in coalified *Trigonocarpus grandis* [9b] or in aerial parts and rhizomes of current species of ferns: *Angiopteris esculenta*, *Cibotium barometz*, *Coniogramme japonica*, *P. aquilinum*, and five species of *Polypodiaceae* [9c-g].

Pteris vittata revealed a VOC fraction very different from that of *P. cretica* with a low diversity (ten volatile compounds), barely any carotenoid derivatives and a majority of shikimic derivatives (69.4%) with the three major odorant benzaldehyde (26%; bitter almond scent), benzyl alcohol (22%; floral notes) and phenylethanal (11%; sweet odor of hyacinth-type). These three VOCs were also detected in five others ferns of the same family analysed in this work but in lower amounts. Lipidic derivatives were represented by nonanal (19.8%; floral-waxy note, [7a]) and 1-octen-3-ol (9.5%; fungal aroma, [3b-d]) recently reported as antifungal agent [10a] and attractant for *Anopheles* and *Aedes* mosquitoes, repellent to *Culex quinquefasciatus* [10b], respectively.

This paper demonstrates that *Peridaceae* can generate a broad spectrum of VOCs for both odorous and bioactive ingredients. Within the former, lipidic derivatives, terpenic compounds and ionone derivatives with fruity odor, herbal scent or floral notes, are the main fragrant components required for cosmetic and hygiene products industries as well as aroma applications: it should be noted that surprising high amount of furan derivatives with caramel or roasty flavor was detected to be used as food additives. Within the last, coumarin derivatives are of various biological interests for pharmaceutical industry and in plant protection products. *Pteridaceae* species resources are potential candidates for bioactive aroma ingredients and for the discovery of new drugs with various therapeutic applications due to their potential anti-inflammatory [10c] and antitumor [10d] promoting properties.

Experimental

Plant material: Fresh aerial parts of ferns were collected in France, as follows: *Pteris cretica*, *P. vittata* and *Adiantum peruvianum*: 31/08/2010, Botanical Garden of Strasbourg; *Cryptogramma crispa*: 01/09/2010, Botanical Garden of Nancy; *Adiantum pedatum*: 01/09/2010, Botanical Garden of Col de Saverne; *Anogramma leptophylla*: 14/04/2010, Le Lavandou (Var);

Cheilanthes maderensis: 13/04/2010, Rayol-Canadel-sur-Mer (Var); Voucher specimens are deposited at the Laboratory of Botany (Faculty of Pharmacy, Limoges, France).

Plant part and GC-MS analyses: Fresh aerial parts of ferns were cubed and extracted with diethyl ether (Carlo Erba, 6 ppm BHT). After one week of maceration at room temperature, the concentrated organic extracts were used for Gas Chromatography Mass Spectrometry (GC-MS) analyses as reported in the literature [5a-c]. The main volatile components of *Pteridaceae* were identified by comparison with National Institute of Standards and Technology Mass Spectral Library [11a-b]. Internal standards (*n*-alkanes) were used as reference points in the calculation of relative retention indices. GC-MS analyses were performed at the « Plateforme d'Analyses Chimiques en Ecologie », technical facilities of the LabEx CeMEB (Centre Méditerranéen pour l'Environnement et la Biodiversité).

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References

- [1] (a) Prelli R, Boudrie M. (2001) *Les fougères et plantes alliées de France et d'Europe occidentale*, Belin, Paris, 1-432; (b) Schuettpelz E, Schneider H, Huiet L, Windham MD, Pryer KM. (2007) A molecular phylogeny of the fern family *Pteridaceae*: Assessing overall relationships and the affinities of previously unsampled genera. *Molecular Phylogenetics and Evolution*, **44**, 1172-1185; (c) Mannar Mannan M, Maridass M, Victor B. (2008) A review on the potential uses of Ferns. *Ethnobotanical Leaflets*, **12**, 281-285; (d) Benniamin A. (2011) Medicinal ferns of North Eastern India with special reference to Anapurna Pradesh. *Indian Journal of Traditional Knowledge*, **10**, 516-522; (e) Maridass M, Raju G. (2010) Conservation status of Pteridophytes, Western Ghats, South India. *Indian Journal of Biotechnology*, **1** (Special Issue), 42-57; (f) May LW. (1978) The economic uses and associated folklore of Ferns and Fern Allies. *The Botanical Review*, **44**, 491-528.
- [2] (a) Limaye AS, Deore GB, Shinde BM, Laware SL. (2010) Assessment of *Adiantum trapeziforme* L. for antioxidant activities. *Asian Journal Experimental Biological Sciences, Special Vol 1*, 79-84; (b) Ghorpade PN, Thakar SB, Dongare MM, Kale MV, Jadhav JP. (2016) Potential of antioxidant capacity and phenol content in four *Cheilanthes* species from Northern Western Ghats. *Asian Journal of Pharmaceutical and Clinical Research*, **9**, 378-382; (c) Singh M, Govindarajan R, Rawat AKS, Khare PB. (2008) Antimicrobial flavonoid rutin from *Pteris vittata* L. against pathogenic gastrointestinal microflora. *American Fern Journal*, **98**, 98-103; (d) De Britto AJ, Gracelin DHS, Kumar PBJR. (2012) *Pteris biaurita* L.: A potential antibacterial fern against *Xanthomonas* and *Aeromonas* bacteria. *Journal of Pharmacy Research*, **5**, 678-680; (e) Paul T, Das B, Apte KG, Banerjee S, Saxena RC. (2012) Hypoglycemic activity of *Pteris vittata* L., a fern on alloxan induced diabetic rats. *Inventi Rapid: Planta Activa*, **2**, 88-91; (f) Gong XL, Chen ZH, Liang NC (2007) Advances in study on chemical constituents and pharmacological activities of plants of genus *Pteris*. *China Journal of Chinese Materia Medica*, **32**, 1382-1387; (g) Pham HN, Michalet S, Bodillis J, Nguyen TD, Nguyen TKO, Le TPQ, Haddad M, Nazaret S, Dijoux-Franca MG (2017) Impact of metal stress on the production of secondary metabolites in *Pteris vittata* L. and associated rhizosphere bacterial communities. *Environmental Science and Pollution Research*, **24**, 16735-16750; (h) Li JH, He CW, Liang NC, Mo LE, Zhang X. (1999) Effects of antitumor compounds isolated from *Pteris semipinnata* L. on DNA topoisomerases and cell cycle of HL- 60 cells. *Zhongguo Yao Li Xue Bao*, **20**, 541-545; (i) Patil SD, Chaudhari MA, Sapkale PV, Chaudhari RB (2013) A recent review on anticancer herbal drugs. *Journal of Drug Discovery and Therapeutics*, **1**, 77-84; (j) Wu MJ, Weng CY, Wang L, Lian TW. (2005) Immunomodulatory mechanism of the aqueous extract of sword brake fern (*Pteris ensiformis* Burm.). *Journal of Ethnopharmacology*, **98**, 73-81.
- [3] (a) Jones DL. (1987) *Encyclopedia of ferns*. Lothian, Port Melbourne, 1-433; (b) Fons F, Rapior S, Eyssartier G, Bessière JM. (2003) Volatile compounds in the *Cantharellus*, *Craterellus* and *Hydnum* genera. *Cryptogamie, Mycologie*, **24**, 367-376; (c) Rapior S, Fons F, Bessière JM. (2003) Volatile flavor constituents of *Lepista nebularis* (clouded Clitocybe). *Cryptogamie, Mycologie*, **24**, 159-166; (d) Fons F, Rapior S, Fruchier A, Saviuc P, Bessière JM. (2006) Volatile composition of *Clitocybe amoenolens*, *Tricholoma caligatum* and *Hebeloma radicosum*. *Cryptogamie, Mycologie*, **27**, 45-55; (e) Fons F, Rapior S, Gargadennec A, Andary C, Bessière JM. (1998) Volatile components of *Plantago lanceolata*. *Acta botanica Gallica*, **145**, 265-269; (f) Maggi F, Papa F, Vittori S. (2012) Gas chromatography for the characterization of the mushroom-like flavor in *Melittis melissophyllum* L. (*Lamiaceae*). *Journal of Essential Oil Research*, **24**, 321-337; (g) Froissard D, Fons F, Bessière JM, Fruchier A, Buatois B, Rapior S. (2014) Volatile organic compounds of six French *Dryopteris* species: Natural odorous and bioactive resources. *Natural Product Communications*, **6**, 1723-1726; (h) Fons F, Froissard D, Bessière JM, Fruchier A, Buatois B, Rapior S. (2013) Fougères et parfumerie. *Annales de la Société d'Horticulture et d'Histoire Naturelle de l'Hérault*, **153**, 96-108; (i) Asakawa Y, Nii K, Higuchi M. (2015) Identification of sesquiterpene lactones in the Bryophyta (mosses) *Takakia*: *Takakia* species are closely related chemically to the Marchantiophyta (liverworts). *Natural Product Communications*, **10**, 5-8; (j) Asakawa Y, Ludwiczuk A, Nagashima F. (2013) Chemical constituents of Bryophytes. Bio- and chemical diversity, biological activity, and chemosystematics. In *Progress in the Chemistry of Organic Natural Products*. Vol. **95**, Kinghorn AD, Falk H, Kobayashi J. (Eds.). Springer, Vienna, 1-796.
- [4] (a) The Good Scents Company (2017) <http://www.thegoodscentscompany.com> (2017/07/18); (b) PubChem, Open Chemistry Database (2017) <https://pubchem.ncbi.nlm.nih.gov/compound> (2017/07/18); (c) Xie L, Chen X. (2017) Mosquito attractant for controlling *Aedes albopictus* and *Culex quinquefasciatus*. *Faming Zhuanli Shenqing*, CN 106305773 A 20170111, 1-14; (d) Kline DL. (1994) Olfactory attractants for mosquito surveillance and control: 1-octen-3-ol. *Journal of the American Mosquito Control Association*, **10**, 280-287; (e) Aprotosoae AC, Hancianu M, Costache II, Miron A. (2014) Linalool: a review on a key odorant molecule with valuable biological properties. *Flavour and Fragrance Journal*, **29**, 193-219; (f) Metoui N, Gargouri S, Amri I, Fezzani T, Jamoussi B, Hamrouni L. (2015) Activity antifungal of the essential oils; aqueous and ethanol extracts from *Citrus aurantium* L. *Natural Product Research*, **29**, 2238-2241; (g) Marquez FM, Moll CN, Herrera PM, Galindo PP, Yuste MM, Sanchez JM. (2004) Activity of the monoterpene derivatives carvacrol, linalool and α -terpineol, Obtained from Aromatic Plants, on *Leishmania infantum*. Multidisciplinarity for Parasites, Vectors and Parasitic Diseases, Proceedings of the European Multicolloquium of

- Parasitology, 9th, Valencia, Spain, July 18-23, 2004, Ed. Mas-Coma, Santiago, 2, 93-96; (h) Müller GC, Junnila A, Butler J, Kravchenko VD, Revay EE, Weiss RW, Schlein Y. (2009) Efficacy of the botanical repellents geraniol, linalool, and citronella against mosquitoes. *Journal of Vector Ecology*, 34, 2-8; (i) Kamatou GPP, Viljoen A. (2008) Linalool – a review of a biologically active compound of commercial importance. *Natural Product Communications*, 3, 1183-1192; (j) The Pherobase (2017) <http://www.pherobase.com/database/kovats/kovats-detail-E,E-pseudoionone.php> (2017/07/18).
- [5] (a) Fons F, Froissard D, Bessièrè JM, Buatois B, Rapior S. (2010) Biodiversity of volatile organic compounds from five French ferns. *Natural Product Communications*, 5, 1655-1658; (b) Froissard D, Fons F, Bessièrè JM, Buatois B, Rapior S. (2011) Volatiles of French ferns and “fougère” scent in perfumery. *Natural Product Communications*, 6, 1723-1726; (c) Fons F, Froissard D, Bessièrè JM, Fruchier A, Buatois B, Rapior S. (2013) Volatile composition of six horsetails: prospects and perspectives. *Natural Product Communications*, 8, 509-512; (d) Gildemeister E. (1913) *The Volatile Oils*, Vol I. John Wiley & Sons, New York, 1-712.
- [6] (a) Hammerschmidt R. (2012) Secondary metabolites and defense: The story continues. *Physiological and Molecular Plant Pathology*, 80, iii-iv; (b) De Girolamo A, Solfrizzo M, Vittì C, Visconti A. (2004) Occurrence of 6-methoxymellein in fresh and processed carrots and relevant effect of storage and processing. *Journal of Agricultural and Food Chemistry*, 52, 6478-6484; (c) Krammer M, Bufler G, Ulrich D, Leitenberger M, Conrad J, Carle R, Kammerer DR. (2012) Effect of ethylene and 1-methylcyclopropene on bitter compounds in carrots (*Daucus carota* L.). *Postharvest Biology and Technology*, 73, 28-36; (d) Magid AA, Voutquenne-Nazabadioko L, Moroy G, Moretti C, Lavaud C. (2007) Dihydroisocoumarin glucosides from stem bark of *Caryocar glabrum*. *Phytochemistry*, 68, 2439-2443; (e) Krohn K, Bahramsari R, Flörke U, Ludewig K, Kliche-Spory C, Michel A, Aust HJ, Draeger S, Schulz B, Antus S. (1997) Dihydroisocoumarins from fungi: isolation, structure elucidation, circular dichroism and biological activity. *Phytochemistry*, 45, 313-320.
- [7] (a) Arctander S. (1969) *Perfume and Flavor Chemicals*. Vol I (A-J) and Vol II (K-Z). Steffen Arctander Publisher, Elizabeth, USA; (b) FEMA (Flavour and Extract Manufacturer Association) <http://www.femaflavor.org/flavor/library/2-methoxy-4-vinylphenol>.
- [8] (a) Lasekan O. (2013) Volatile constituents of roasted tigernut oil (*Cyperus esculentus* L.). *Journal of the Science of Food and Agriculture*, 93, 1055-1061. (b) Zhao YP, Wang L, Li JM, Pei GR, Liu QS. (2011) Comparison of volatile compounds in two brandies using HS-SPME coupled with GC-O, GC-MS and Sensory evaluation. *South African Journal of Enology & Viticulture*, 32, 9-20; (c) Sigma Aldrich, Ingredients Catalog: Flavors and Fragrances (http://www.sigmaaldrich.com/ifb/fnf_2014); (d) Merck & Co., Inc. (1996) *The Merck Index, An Encyclopedia of Chemicals, Drugs, and Biologicals*, 12th Ed. Merck and Co., Inc., Whitehouse Station, NJ, 1-1741; (e) Abraham K, Gürtler R, Berg K, Heinemeyer G, Lampen A, Appel KE. (2011) Toxicology and risk assessment of 5-hydroxymethylfurfural in food. *Molecular Nutrition & Food Research*, 55, 667-678; (f) Amanpour A, Selli S. (2016) Differentiation of volatile profiles and odor activity values of turkish coffee and French press coffee. *Journal of Food Processing and Preservation*, 40, 1116-1124; (g) Perestrelo R, Silva CL, Câmara JS. (2015) Quantification of furanic derivatives in fortified wines by a highly sensitive and ultrafast analytical strategy based on digitally controlled microextraction by packed sorbent combined with ultrahigh pressure liquid chromatography. *Journal of Chromatography A*, 1381, 54-63; (h) Paravisini L, Gourrat-Pernin K, Gouttefangeas C, Moreton C, Nigay H, Dacremont C, Guichard E. (2012) Identification of compounds responsible for the odorant properties of aromatic caramel. *Flavour and Fragrance Journal*, 27, 424-432.
- [9] (a) Froissard D, Rapior S., Bessièrè JM, Buatois B, Fruchier A, Sol V, Fons F. (2015) *Asplenioideae* species as a reservoir of volatile organic compounds with potential therapeutic properties. *Natural Product Communications*, 10, 1079-1083; (b) Zodrow EL, Helleur R, Werner-Zwanziger U, Chen B, D'Angelo JA. (2013) Spectrochemical study of coalified *Trigonocarpus grandis* (Pennsylvanian tree-fern ovule, Canada): Implications for fossil-organ linkage. *International Journal of Coal Geology*, 109-110, 24-35; (c) Chen Y, Tao Y, Lian X, Wang L, Zhao Y, Jiang J, Zhang Y. (2010) Chemical constituents of *Angiopteris esculenta* including two new natural lactones. *Food Chemistry*, 122, 1173-1175; (d) Cuong NX, Minh CV, Kiem PV, Huong HT, Ban NK, Nhiem NX, Tung NH, Jung JW, Kim HJ, Kim SY, Kim JA, Kim YH. (2009) Inhibitors of osteoclast formation from rhizomes of *Cibotium barometz*. *Journal of Natural Products*, 72, 1673-1677; (e) Fang CW, Chen JJ, Liu SJ. (2010) Studies on the chemical constituents of rhizome of *Coniogramme japonica*. *Zhong Yao Cai*, 33, 557-559; (f) Leach H, Barber GD, Evans IA, Evans WC. (1971) Isolation of an active principle from the bracken fern that is mutagenic, carcinogenic and lethal to mice on intraperitoneal injection. *Proceedings of the Biochemical Society*, 124, 13P-14P; (g) Kessler M, Connor E, Lehnert M. (2015) Volatile organic compounds in the strongly fragrant fern genus *Melpomene* (Polypodiaceae). *Plant Biology*, 17, 430-436.
- [10] (a) Zhang JH, Sun HL, Chen SY, Zeng L, Wang TT. (2017) Anti-fungal activity, mechanism studies on α -phellandrene and nonanal against *Penicillium cyclopium*. *Botanical Studies* 58:13. DOI: 10.1186/s40529-017-0168-8; (b) Xu P, Zhu F, Buss GK, Leal WS. (2015) 1-Octen-3-ol – the attractant that repels. *F1000Research*, 4, 156. DOI: 10.12688/f1000research.6646.1; (c) Chen YC, Tsai WJ, Wu MH, Lin LC, Kuo YC. (2007) Suberosin inhibits proliferation of human peripheral blood mononuclear cells through the modulation of the transcription factors NF-AT and NF-kappaB. *British Journal of Pharmacology*, 150, 298-312; (d) Kim JK, Kim JY, Kim HJ, Park KG, Harris RA, Cho WJ, Lee JT, Lee IK. (2013) Scoparone exerts anti-tumor activity against DU145 prostate cancer cells via inhibition of STAT3 activity. *PLoS One*. Nov 15;8(11):e80391. DOI: 10.1371/journal.pone.0080391. eCollection 2013.
- [11] (a) National Institute of Standard and Technology. (2005) *PC version of the NIST / EPA / NIH Mass Spectra Database*, Gaithersburg, Maryland, USA; (b) Adams RP. (2007) *Identification of Essential Oil Components by Gas Chromatography/Mass Spectroscopy*. 4th Ed., Allured, Carol Stream, IL, USA.

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