NATURAL PRODUCT COMMUNICATIONS

An International Journal for Communications and Reviews Covering all Aspects of Natural Products Research



www.naturalproduct.us



Natural Product Communications

EDITOR-IN-CHIEF

DR. PAWAN K AGRAWAL

Natural Product Inc. 7963, Anderson Park Lane, Westerville, Ohio 43081, USA agrawal@naturalproduct.us

EDITORS

PROFESSOR ALESSANDRA BRACA Dipartimento di Chimica Bioorganicae Biofarmacia, Universita di Pisa, via Bonanno 33, 56126 Pisa, Italy braca@farm.unipi.it

PROFESSOR DEAN GUO State Key Laboratory of Natural and Biomimetic Drugs, School of Pharmaceutical Sciences, Peking University, Beijing 100083, China gda5958@163.com

PROFESSOR YOSHIHIRO MIMAKI

School of Pharmacy, Tokyo University of Pharmacy and Life Sciences, Horinouchi 1432-1, Hachioji, Tokyo 192-0392, Japan mimakiy@ps.toyaku.ac.jp

PROFESSOR STEPHEN G. PYNE

Department of Chemistry University of Wollongong Wollongong, New South Wales, 2522, Australia spyne@uow.edu.au

PROFESSOR MANFRED G. REINECKE

Department of Chemistry, Texas Christian University, Forts Worth, TX 76129, USA m.reinecke@tcu.edu

PROFESSOR WILLIAM N. SETZER

Department of Chemistry The University of Alabama in Huntsville Huntsville, AL 35809, USA wsetzer@chemistry.uah.edu

PROFESSOR YASUHIRO TEZUKA

Institute of Natural Medicine Institute of Natural Medicine, University of Toyama, 2630-Sugitani, Toyama 930-0194, Japan tezuka@inm.u-toyama.ac.jp

PROFESSOR DAVID E. THURSTON Department of Pharmaceutical and Biological Chemistry, The School of Pharmacy, University of London, 29-39 Brunswick Square, London WCIN 1AX, UK david.thurston@pharmacy.ac.uk

HONORARY EDITOR

PROFESSOR GERALD BLUNDEN The School of Pharmacy & Biomedical Sciences, University of Portsmouth, Portsmouth, POI 2DT U.K. axuf64@dsl.pipex.com

ADVISORY BOARD

Prof. Berhanu M. Abegaz Gaborone, Botswana Prof. Viqar Uddin Ahmad Karachi, Pakistan Prof. Øyvind M. Andersen Bergen, Norway Prof. Giovanni Appendino Novara, Italy Prof. Yoshinori Asakawa Tokushima, Japan Prof. Lee Banting Portsmouth, U.K. Prof. Julie Banerji Kolkata, India Prof. Alejandro F. Barrero Granada, Spain Prof. Anna R. Bilia Florence, Italy Prof. Maurizio Bruno Palermo, Italy Prof. César A. N. Catalán Tucumán, Argentina Prof. Josep Coll Barcelona, Spain Prof. Geoffrey Cordell Chicago, IL, USA Prof. Cristina Gracia-Viguera Murcia, Spain Prof. Duvvuru Gunasekar Tirupati, India Prof. A.A. Leslie Gunatilaka Tucson, AZ, USA Prof. Kurt Hostettmann Lausanne, Switzerland Prof. Martin A. Iglesias Arteaga Mexico, D. F, Mexico Prof. Jerzy Jaroszewski Copenhagen, Denmark

Prof. Leopold Jirovetz Vienna, Austria Prof. Karsten Krohn Paderborn, Germany Prof. Hartmut Laatsch Gottingen, Germany Prof. Marie Lacaille-Dubois Dijon, France Prof. Shoei-Sheng Lee Taipei, Taiwan Prof. Francisco Macias Cadiz, Spain Prof. Imre Mathe Szeged, Hungary Prof. Joseph Michael Johannesburg, South Africa Prof. Ermino Murano Trieste, Italy Prof. M. Soledade C. Pedras Saskatoon, Canada Prof. Luc Pieters Antwerp, Belgium Prof Peter Proksch Düsseldorf, Germany Prof Phila Rahariyelomanana Tahiti, French Polynesia Prof. Monique Simmonds Richmond, UK Prof. Valentin Stonik Vladivostok, Russia Prof. Winston F. Tinto Barbados, West Indies Prof. Karen Valant-Vetschera Vienna, Austria Prof. Peter G. Waterman Lismore, Australia

INFORMATION FOR AUTHORS

Full details of how to submit a manuscript for publication in Natural Product Communications are given in Information for Authors on our Web site http://www.naturalproduct.us.

Authors may reproduce/republish portions of their published contribution without seeking permission from NPC, provided that any such republication is accompanied by an acknowledgment (original citation)-Reproduced by permission of Natural Product Communications. Any unauthorized reproduction, transmission or storage may result in either civil or criminal liability.

The publication of each of the articles contained herein is protected by copyright. Except as allowed under national "fair use" laws, copying is not permitted by any means or for any purpose, such as for distribution to any third party (whether by sale, loan, gift, or otherwise); as agent (express or implied) of any third party; for purposes of advertising or promotion; or to create collective or derivative works. Such permission requests, or other inquiries, should be addressed to the Natural Product Inc. (NPI). A photocopy license is available from the NPI for institutional subscribers that need to make multiple copies of single articles for internal study or research purposes.

To Subscribe: Natural Product Communications is a journal published monthly. 2011 subscription price: US\$1,995 (Print, ISSN# 1934-578X); US\$1,995 (Web edition, ISSN# 1555-9475); US\$2,495 (Print + single site online); US\$595 (Personal online). Orders should be addressed to Subscription Department, Natural Product Communications, Natural Product Inc., 7963 Anderson Park Lane, Westerville, Ohio 43081, USA. Subscriptions are renewed on an annual basis. Claims for nonreceipt of issues will be honored if made within three months of publication of the issue. All issues are dispatched by airmail throughout the world, excluding the USA and Canada.

NPC Natural Product Communications

Volatiles of French Ferns and "fougère" Scent in Perfumery

Didier Froissard^a, Françoise Fons^b*, Jean-Marie Bessière^c, Bruno Buatois^c and Sylvie Rapior^b

^aLaboratoire de Botanique, Faculté de Pharmacie de Limoges, 2 rue du Dr Marcland, F-87025 Limoges cedex, France

^bLaboratoire de Botanique, Phytochimie et Mycologie, Faculté de Pharmacie (Université Montpellier 1), UMR 5175 CEFE, B.P. 14 491, 15 avenue Charles Flahault, F-34093 Montpellier cedex 5, France

^cCentre d'Ecologie Fonctionnelle et Evolutive – Plateforme d'analyses chimiques en écologie, UMR 5175 CEFE, 1919 Route de Mende, F-34293 Montpellier cedex 5, France

francoise.fons@univ-montp1.fr

Received: March 23rd, 2011; Accepted: June 24th, 2011

Six French ferns were investigated for volatile organic compounds (VOC) by GC-MS using organic solvent extraction. Seventy-seven VOC biosynthesized from the shikimic, lipidic and terpenic pathways, including isoprenoid derivatives, were identified from these putative natural resources. *Asplenium trichomanes* subsp. *trichomanes* contained mainly polyketides with an oily or waxy odor. (*E*)-2-Hexenal and (*Z*)-3-hexenol, responsible for the "green odor", were found in high contents in *Polystichum setiferum, Dryopteris dilatata* and *Phegopteris connectilis*. In the last, 7.4% of coumarin with a cut hay scent was highlighted from the volatile fraction. (*E*)-3-Hexenoic acid and (*E*)-2-hexenoic acid, both with herbal and fruity notes, were identified in *Gymnocarpium dryopteris* and *Pteridium aquilinum*. 1-Octen-3-ol, well-known for its mushroom-like odor, was abundant in all analyzed French ferns. While the "fougère" fragrance is claimed by the perfumers to be a fantasy scent, coumarin, (*E*)-2-hexenal, (*Z*)-3-hexenol and 1-octen-3-ol are the main odorous components of the perfumes belonging to the fougère accord family. This suggests that the fougère scent from the perfumers' imagination is a natural fragrance.

Keywords: Ferns, fougere, volatile organic compounds, (E)-2-hexenal, (E)-3-hexenoic acid, coumarin, 1-octen-3-ol, Pteridophyta.

Ferns and fern allies (Monilophytes and Lycophytes [1a,1b]) are widely distributed in France due to the geological variety of soils, different climatic interactions, contrasted altitudes and various ecosystems. Indeed, there are over one hundred species of native ferns and fern allies in mainland France [1c]. In this way the French fern flora shows a great biodiversity of volatile organic compounds (VOC) [1d]. These plants have given perfumers great inspiration for a long time. In 1882, the French perfumer Jean-François Houbigant created a modern perfume called "Fougère Royale" with aromatic, spicy, oakmossy notes adding synthetic coumarin for the first time in perfumery [2a-2c]. Nowadays, the French word "fougère" is commonly used in perfume composition. The fougère note refers to a basic scent of great interest for fine fragrances, cosmetics and soaps. In the perfume classification described by the French society of perfumers, fougère is one of the seven scent families contributing to the olfactory description of a fragrance [2d,2e]. However, few reports [1d,3a-3f] have been published on VOC from wild ferns. As a part of our ongoing work on the VOC of Monilophytes [1d], six French ferns were investigated for their volatile profiles using GC-MS: Asplenium trichomanes subsp. trichomanes L. (Aspleniaceae), Drvopteris dilatata (Hoffm.) A. Grav. Polvstichum setiferum (Forskal) Woynar (Dryopteridaceae), Gymnocarpium dryopteris (L.)

Newman (Woodsiaceae), *Pteridium aquilinum* (L.) Kuhn (Dennstaedtiaceae), and *Phegopteris connectilis* (Michx) Watt (Thelypteridaceae). The main volatile components were compared, in particular, with those reported in the literature, including the five French ferns previously studied [1d].

Aerial parts of fresh ferns were analyzed for volatizable and volatile organic compounds. Seventy-seven components biosynthesized from the shikimic, lipidic and terpenic pathways were identified from the concentrated organic extracts of the six ferns (Table 1).

Twenty-eight volatiles were identified for *A. trichomanes* subsp. *trichomanes*, with mostly polyketide compounds (87.5%) and minor aromatic compounds (8.2%), i.e., benzofuran with an aromatic note [4a]. High amounts of fatty acid derivatives (alcohols, aldehydes, acids) from C_7 to C_{10} series, i.e., (*E*)-2-heptenal (green fatty note), nonanal, octanoic acid (fatty, soapy odor), (*E*)-2-decenal, (*E*)-2-decenol (waxy note), nonanoic acid (waxy dairy note), and decadienal isomers were found [4b-4d]. This fern volatile composition showed a likeness to the *Adiantum capillus-veneris* volatiles [1d]: lack of green odor C₆-compounds, mainly heavy polyketides with plastic or fatty odor (decadienal isomers, (*E*)-2-decenal).

	Table 1: Percentage of volatile organic compounds ^a in fresh aerial part of ferns.							
Compounds	RI ^b	Asplenium	Dryopteris	Polystichum	Gymnocarpium	Pteridium	Phegopteris	
Aromatic compounds		trichomanes	dilatata 13.9	setiferum	dryopteris	aquilinum	connectilis	
Benzaldehyde	962	0.2	2.2	1.9	0.8	2.3	23.3	
Phenylethanal	1046				1.7		5.3	
Acetophenone 2 Phonylathanal	1068	0.1	4.0	0.9			0.8	
Benzoic acid	1187	0.1	1.4	1.6	3.4	5.2		
Benzothiazole	1228		0.6		0.6			
Benzofuran	1235	7.4						
4-Amino-2-methoxyphenol	1243		1.6	1.5	0.8	0.0		
Hvdroquinone	1283		1.0		0.8	0.9	1.5	
4-Vinylguaiacol	1318					2.6	1.9	
4-Hydroxybenzaldehyde	1388	0.7						
Isovanillin	1404		0.8	0.9		1.4	7.4	
Methyl 4-hydroxybenzoate	1431				3.4		7.4	
Methyl vanillate	1518			0.6	3.4			
4-Hydroxybenzoic acid	1551			3.2	0.6		0.6	
Vanillic acid	1592		1.0			4.3	5.9	
Coniferol	1639						0.6	
Polyketide compounds	1752	87.5	55.4	61.9	26.7	63.0	50.6	
Isovaleric acid	850	0.4			1.4	1.7		
(<i>E</i>)-2-Hexenal	851		10.1	6.6	2.0	2.2	9.1	
(L)-3-mexenon Furfurol	855 863		3.0	3.2	1./	2.5 1.4	3.2	
(E)-2-Hexenol	866		2.8			1.7		
Hexanol	870					1.4		
Heptanal	902	0.6	1.6			1.7	2.1	
Sorbaldehyde	914			1.6		2.0		
(E)-2-Hentenal	956	8.0		0.9		2.0	4 2	
1-Octen-3-one	977	0.4	0.8	1.5	0.6		1.1	
1-Octen-3-ol	983	3.4	13.1	21.3	4.5	9.8	6.8	
Octan-2,3-dione	985	0.4	0.6	1.3	0.8		1.3	
2-Pentylluran (2F 47)-Heptadienal	990	0.4			11			
3-Octanol	1000	0.1	2.0	1.5				
Hexanoic acid	1009	1.2	2.0	1.1	1.4	2.3		
(2E, 4E)-Heptadienal	1013	1.2	2.4	6.4	7.1	4.2	2.7	
(E)-3-Hexenoic acid	1017		5.4 5.0	0.4	/.1	4.3	2.7	
(E)-2-Octenal	1055	1.0	5.0	1.9	2.0	1.2		
(E)-2-Octenol	1072	0.7	1.4	2.2			0.6	
1-Octanol	1089	0.9					1.3	
Nonanal	1106	4.3	4.8	3.4		17	4.6	
3 5-Dihydroxy-2 3-dihydro-4-pyranone	1140					10.1		
(<i>E</i>)-2-Nonenal	1162		0.8	1.7			1.9	
(E)-2-Nonenol	1172			1.3				
Octanoic acid	1193	18.4				5.5		
(E)-2-Decenal	1220	2.1				5.5		
(E)-2-Decenol	1263	9.2					4.6	
4-Methyl-3-vinylmaleimide	1273	10.0		1.6		0.9		
Nonanoic acid $(2F 47)$ Decadienal	1283	12.3	1.2		1.1		1.1	
(2E,4E)-Decadienal	1321	4.7		1.3			1.5	
(E)-2-Undecenal	1366	0.6		3.2				
9-Oxononanoic acid	1498	2.8				5.0		
Tridec-9-enoic acid	1060	9.8 1.2	2.6		2.0	5.8 4.6	3.0	
Monoterpenic compounds	1//3	0	0	1.3	4.0	4.0 0	1.5	
α-Pinene	933			1.3				
trans-Linalool oxide (pyran)	1393				3.4		1.5	
cis-Linalool oxide (pyran)	1396	13	30.6	26.2	0.6	20.2	22.6	
NI ^c	1582	4.5	2.6	20.2	2.8	20.2	22.0	
NI ^c	1590				3.1			
NI ^e	1597	0.6	2.8		7.4	1.4	3.2	
5-Hydroxy-5,6-dihydro-β-ionol NI°	1606		1.0		1 1		0.6	
4-Hydroxy-7 8-dihydro-8-ionone	1631		2.0		1.1		2.5	
6-Hydroxy-α-ionone	1635		2.2	3.4	2.0	1.7	2.0	
3-Oxo-α-ionol	1644	0.4	6.0		1.1			
4-Oxo-7,8-dihydro-β-ionone	1649			2.1	3.7			
^{1N1} 3-Ωxo-α-ionone	1649			2.1	6.5	17	0.6	
5-Hydroxy-5,6,7,8-tetrahydroionone	1652			2.5	6.5	1.7	0.0	
4-Hydroxy-5,6-epoxy-β-ionol	1661	3.4	5.8			2.3	12.7	
4-Oxo-5,6-epoxy-β-ionol	1673			1.5	13.6		2.1	
4-UX0-/,δ-ainydro-β-ionone 2-Hydroxy-5 6-epoxy-β-ionone	16/8			1.5	2.8		2.1	
3-Hydroxy-5,6-epoxy-B-ionone	1691		5.8	13.9		2.3	0.8	
3-Oxo-7,8-dihydro-α-ionol	1705					5.2		
3-Oxo-7,8-dihydro-β-ionol	1721			3.0		2.9		
<u>3-Ux0-6-hydroxy-α-ionone</u>	1800	2.95/20	17.56 / 150	7.05 /100	2.59 / 50	0.9	19.9/200	

Extraction process
2.83 / 30
^a Relative percentage of the VOC based on the GC-MS chromatographic area
^b R1 = Retention Indices on SLBTM-5MS column (Supelco)
^c NI = non identified
^d Fresh Weight of fern and volume of diethyl ether used for the extraction (g/mL).

Thirty compounds participated in the general aroma of *D. dilatata* with aromatic compounds (13.9%) such as 2-phenylethanol, with a rose odor [4e,4f] and benzaldehyde with an almond note [4g,4h] already found in *Blechnum spicant* [1d]. Polyketides formed the major group of compounds (55.4%) with (*E*)-2-hexenal (10.1%), 1-octen-3-ol (13.1%), and (*E*)-2-hexenoic acid (5%), with a powerful fruity, herbal, and fatty odor, and nonanal (4.8%) with an orange and green scent [4i].

The broad spectrum of volatile components (thirty-one) identified in *P. setiferum* contributed to its complex smell, including, among others, polyketide derivatives (61.9% of the volatile fraction), isoprenoid derivatives (26.2%), various aromatics (10.6%) and an odorous monoterpene, α -pinene. The main volatiles were 1-octen-3-ol (21.3%), (E)-2-hexenal (6.6%) and (E)-3-hexenoic acid (6.4%), with a honey odor that is somewhat waxy, fruity and herbal [4b]. The first, the C₈-alcohol well-known for its mushroom odor [4e,4g,4i,4j], was abundant in the eleven ferns analyzed in our studies [1d]. High contents of (E)-2hexenal, (Z)-3-hexenol and C₆-acid derivatives were also measured in D. dilatata (13.1%), P. connectilis (12.3%) and the previously reported Athyrium filix-femina (15.1%) [1d]. It should be noted that C_6 -aldehydes and -alcohols are commonly released after either plant stress or herbivory-induced damage [5a-5b]. The C₆-derivatives are notably responsible for the green odor of the plants [4]. This pleasant odor would be beneficial for mammals, with an antidepressant-like effect [5c].

The volatile pattern (twenty-nine volatiles) of G. drvopteris was dominated by a large number of isoprenoid derivatives (50.7%), i.e., ionone derivatives. The investigation indicated lower amounts of polyketides and aromatics (26.7% and 14,7%, respectively), as well as 1octen-3-ol (4.5%) and (E)-3-hexenoic acid (7.1%). The terpenic pathway produces either fragrant monoterpenes (α -terpineol, pinenes) or sesquiterpenes (β -caryophyllene, (E)-nerolidol), which give a pleasant scent [4f,4h] to the concerned fern, for example, Oreopteris limbosperma [1d] and Drvopteris filix-mas, while sesquiterpene glucosides, such as the illudane-type ptaquiloside may induce a carcinogenic effect, as in the case of *P. aquilinum* [6a-6b]. Glycosides of hemiterpenes, diterpenes, sesterpenes or triterpenes have been isolated from different ferns, as well as carotenoids [6c-6f], but are not accessible by GC-MS. Only ionone derivatives, produced after carotenoid degradation and widely found in six of the eleven studied ferns, were detectable by GC-MS.

The complex volatile profile of *P. aquilinum* is based on twenty-nine volatiles, mainly polyketides (63%), as well as isoprenoids (20.2%) and aromatics (16.8%). Among the former group, 3,5-dihydroxy-2,3-dihydro-4-pyranone (10.1%), 1-octen-3-ol (9.8%), (*E*)-2-hexenoic acid (7.2%) and 5-hydroxymethylfurfural (5.5%) were the major components. Table 1 lists also benzoic acid and oxo-7,8-

dihydro- α -ionol (5.2% each), as well as vanillic acid and 4-vinylguaiacol, with a spicy, clove, smoky odor [4b,4k,4l].

Thirty-two volatiles were identified in *P. connectilis*, mainly polyketides (50.6%), aromatics (25.3%) and isoprenoids (22.6%). Coumarin (hay and dried herb odor; 7.4%), 1-octen-3-ol (6.8%), vanillic acid (sweet cream with vanilla aroma; 5.9%), phenylethanal (5.3% hyacinth, lilac note), nonanal (4.6%) and (*E*)-2-decenol (4.6%) were the main volatile odorous components [4h-4j,4m].

While the fougère fragrance is claimed by the perfumers to be a fantasy scent [2a,2b], coumarin (hay and dried herb odor), (*E*)-2-hexenal and (*Z*)-3-hexenol (green odors), 1octen-3-ol (mushroom-like odor), terpenic compounds with citrus, floral and spicy notes (α -terpineol, (*E*)nerolidol, β -caryophyllene, linalool, pinenes, limonene, γ terpinen-7-al, ionone derivatives...) are the main fragrant components of the perfumes belonging to the fougère accord family. Most of these sweet-smelling compounds have been found in the wild French ferns [1d]. Because of the increasing requirements of the perfumery, cosmetic and hygienic product industries for natural odor materials, fern resources are of great interest as natural sources of odorous ingredients.

Experimental

Plant material: Fresh aerial parts of ferns were collected from metropolitan France as follows: *Asplenium trichomanes* subsp. *trichomanes* L., *Dryopteris dilatata* (Hoffm.) A. Gray, *Gymnocarpium dryopteris* (L.) Newman and *Pteridium aquilinum* (L.) Kuhn: 20/9/2009, Meymac, Corrèze. *Polystichum setiferum* (Forskål) Woynar: 14/09/2009, Botanical Garden of Limoges. *Phegopteris connectilis* (Michx) Watt: 14/7/2009, Gimelles-Cascades, Corrèze. Voucher specimens are deposited at the Laboratory of Botany (Faculty of Pharmacy, Limoges, France).

Plant part and GC-MS analyses: Aerial parts of fresh ferns were cubed and extracted with diethyl ether (Carlo Erba, 6 ppm BHT; Table 1). After maceration for one week at room temperature, the concentrated organic extracts were used for Gas Chromatography-Mass Spectrometry (GC-MS) analyses, as previously described [1d]. VOC were identified by comparison with National Institute of Standards and Technology Mass Spectral Library [7a], retention indices reported in the literature [7b] and our own data bank.

Acknowledgments - The authors are indebted to the Botanical Garden of Limoges for providing *Polystichum setiferum*. We are also grateful to Professor Alain Fruchier (ENSCM, Montpellier, France) for his helpful comments on chemistry.

References

- (a) Pryer KM, Schuettpelz E, Wolf PG, Schneider H, Smith AR, Cranfill R. (2004) Phylogeny and evolution of ferns (Monilophytes) with a focus on the early leptosporangiate divergences. *American Journal of Botany*, 91, 1582-1598; (b) Prelli R. (2010) La classification phylogénétique des ptéridophytes actuelles. *Le Journal de Botanique de la Société Botanique de France*, 49, 37-47; (c) Prelli R, Boudry M. (2001) *Les fougères et plantes alliées de France et d'Europe occidentale*, Berlin, Paris, 1-432; (d) Fons F, Froissard D, Bessière JM, Buatois B, Rapior S. (2010) Biodiversity of volatile organic compounds from five French ferns. *Natural Product Communications*, 5, 1655-1658.
- (a) Anonis DP. (1994) Fougère in fine fragrances. *Perfumer & Flavorist 19* (1), 1-5; (b) Anonis DP. (1994) Fougère in colognes, cosmetics and soaps, and in men's fragrances. *Perfumer & Flavorist 19* (3), 35-39; (c) Parfums Houbigant Paris (2011) http://www.houbigant-parfum.com/main.html; (d) Société Française des Parfumeurs (2011) La classification des parfums http://www.parfumeur-createur.com/rubrique.php3?id_rubrique=20; (e) de Nicolaï P. (2008) A smelling trip in the past: the influence of synthetic materials on the history of perfumery. *Chemistry & Biodiversity*, 5, 1137-1146.
- (a) Briggs LH, Sutherland MD. (1947) A terpene-type essential oil from a fern (*Paesia scaberula*). Nature, 160, 333; (b) Juliani HR, Zygadlo JA, Scrivanti R, de la Sota E, Simon JE. (2004) The essential oil of Anemia tomentosa (Savigny) Sw. var. anthriscifolia (Schrad.) Mickel. Flavour and Fragrance Journal, 19, 541-543; (c) Cheng CG, Mao JY. (2005) Constitution of volatile oils from three kinds of pteridophyte plants. Linchan Huaxue Yu Gongye Chemical and Industry of Forest Products, 25, 107-110; (d) Naseri NG, Ashnagar A, Nia SJ. (2006) Isolation and structural determination of the major chemical compounds possibly found in the leaves of maidenhair plant (Adiantum capillus-veneris L.) grown around the city of Dezful, Iran. International Journal of Chemical Sciences, 4, 874-880; (e) Miyazawa M, Horiushi E, Kawata J. (2007) Components of the essential oil from Matteuccia struthiopteris. Journal of Oleo Science, 56, 457-461; (f) Imbiscuso G, Trotta A, Maffei M, Bossi S. (2009) Herbivory induces a ROS burst and the release of volatile organic compounds in the fern Pteris vittata L. Journal of Plant Interactions, 4, 15-22.
- (a) Merck & Co., Inc. (1996) The Merck Index, 12th Ed., Whitehouse Station, NJ, USA. 1-1741; (b) The Good Scents Company [4] (2011) http://www.thegoodscentscompany.com/; (c) Rapior S, Fons F, Bessière JM. (2000) The fenugreek odor of Lactarius helvus. Mycologia, 92, 305-308; (d) Tahrouch S, Rapior S, Belahsen Y, Bessière JM, Andary C. (1998) Volatile constituents of Peganum harmala. Acta botanica Gallica, 145, 121-124; (e) Rapior S, Konska G, Guillot J, Andary C, Bessière JM. (2000) Volatile composition of Laetiporus sulphureus. Cryptogamie, Mycologie, 21, 67-72; (f) Rapior S, Fons F, Bessière JM. (2003) Volatile flavor constituents of Lepista nebularis (Clouded Clitocybe). Cryptogamie, Mycologie, 24, 159-166; (g) Rapior S, Breheret S, Talou T, Pélissier Y, Bessière JM. (2002) The anise-like odor of Clitocybe odora, Lentinellus cochleatus and Agaricus essettei. Mycologia, 94, 373-376; (h) Fons F, Rapior S, Fruchier A, Saviuc P, Bessière JM. (2006) Volatile composition of Clitocybe amoenolens, Tricholoma caligatum and Hebeloma radicosum. Cryptogamie, Mycologie, 27, 45-55; (i) Fons F, Rapior S, Evssartier G, Bessière JM. (2003) Volatile compounds in the Cantharellus, Craterellus and Hydnum genera. Cryptogamie, Mycologie, 24, 367-376; (j) Fons F, Rapior S, Gargadennec A, Andary C, Bessière JM. (1998) Volatile components of Plantago lanceolata (Plantaginaceae). Acta botanica Gallica, 145, 265-269; (k) Coghe S, Benoot K, Delvaux F, Vanderhaegen B, Delvaux FR. (2004) Ferulic acid release and 4-vinylguaicol formation during brewing and fermentation: indications for feruloyl esterase activity in Saccharomyces cerevisiae. Journal of Agricultural and Food Chemistry, 52, 602-608; (1) Komes D, Ulrich D, Lovric T. (2006) Characterization of odor-active compounds in Croatian Rhine Riesling wine, subregion Zagorje. European Food Research Technology, 222, 1-7; (m) Chiron N, Michelot D. (2005) Odeur des champignons : chimie et rôle dans les interactions biotiques -Une revue. Cryptogamie, Mycologie, 26, 299-364.
- [5] (a) Hatanaka A. (1993) The biogeneration of green odour by green leaves. *Phytochemistry*, 34, 1201-1218; (b) Shimoda T. (2010) A key volatile infochemical that elicits a strong olfactory response of the predatory mite *Neoseiulus californicus*, an important natural enemy of the two-spotted spider mite *Tetranychus urticae*. *Experimental Applied Acarology*, 50, 9-22; (c) Fujita S, Ueki S, Miyoshi M, Watanabe T. (2010) "Green odor" in inhalation by stressed rat dams reduces behavioral and neuroendocrine signs of prenatal stress in the offspring. *Hormones and Behavior*, 58, 264-272.
- (a) Potter DM, Baird MS. (2000) Carcinogenic effects of ptaquiloside in bracken fern and related compounds. British Journal of Cancer, 83, 914-920; (b) Jensen PH, Jacobsen AS, Hansen HCB, Juhler RK. (2008) Quantification of ptaquiloside and pterosin B in soil and groundwater using LC-MS/MS. Journal of Agricultural and Food Chemistry, 56, 9848-9854; (c) Toyota M, Oiso Y, Asakawa Y. (2002) New glycosides from the Japanese fern Hymenophyllum barbatum. Chemical and Pharmaceutical Bulletin, 50, 508-514; (d) Murakami T, Saiki Y. (1989) Chemosystematics of di- and sesquiterpenoids in polypodiaceous ferns. Biochemical Systematics and Ecology, 17, 131-140; (e) Imperato F. (2002) Recent advances in isoprenoid compounds from Pteridophyta. Current Topics in Phytochemistry, 5, 153-223; (f) Czeczuga B. (1985) Carotenoids in sixty-six representatives of the Pteridophyta. Biochemical Systematics and Ecology, 13, 221-230.
- [7] (a) National Institute of Standard and Technology (2005) *PC version of the NIST / EPA / NIH Mass Spectral Database*, Gaithersburg, Maryland, USA; (b) Adams RP. (2007) *Identification of Essential Oil Components by Gas Chromatography / Mass Spectroscopy*. 4th edition, Allured, Carol Stream, IL, USA.

Effects of pH, Sample Size, and Solvent Partitioning on Recovery of Soluble Phenolic Acids and Isoflavonoids in Leaves and Stems of Red Clover (<i>Trifolium pratense</i> cv. Kenland)				
Isabelle A. Kagan	1657			
Arbutin Derivatives from the Seeds of <i>Madhuca latifolia</i> Shazia Khan, M. Nadeem Kardar and Bina S. Siddiqui				
Quinic Acids from <i>Aster caucasicus</i> and from Transgenic Callus Expressing a β-Amyrin Synthase Paola Pecchia, Maria Cammareri, Nicola Malafronte, M. Federica Consiglio, Maria Josefina Gualtieri and Clara Conicella	1665			
Cytotoxic Activity and Cell Cycle Analysis of Hexahydro-curcumin on SW 480 Human Colorectal Cancer Cells Chung-Yi Chen, Woei-Ling Yang and Soong-Yu Kuo	1671			
Does the Combination of Resveratrol with Al (III) and Zn (II) Improve its Antioxidant Activity? Karina Dias and Sofia Nikolaou	1673			
Photosensitization Mechanisms of Triplet Excited Stateβ-Lapachone. A Density Functional Theory Study Liang Shen	1677			
AFLP Marking and Polymorphism among Progenies of <i>Gymnema sylvestre</i> : an Important Medicinal Plant of India Magda Abbaker Osman, Sunita Singh Dhawan, Janak Raj Bahl, Mahendra P Darokar and Suman P S Khanuja	1679			
Antioxidant Activity of Protein Hydrolysates from Aqueous Extract of Velvet Antler (<i>Cervus elaphus</i>) as Influenced by Molecular Weight and Enzymes				
Lei Zhao, Yang-Chao Luo, Cheng-Tao Wang and Bao-Ping Ji	1683			
Effects of <i>Sideritis euboea</i> (Lamiaceae) Aqueous Extract on IL-6, OPG and RANKL Secretion by Osteoblasts Eva Kassi, Anna Paliogianni, Ismene Dontas, Nektarios Aligiannis, Maria Halabalaki, Zoi Papoutsi, Alaxios Leandros Skaltsounis and Paraskavi Mouteatoou	1680			
In vitro Antiprotozoal Activity of Extracts of five Turkish Lamiaceae Species	1007			
Hasan Kirmizibekmez, Irem Atay, Marcel Kaiser, Erdem Yesilada and Deniz Tasdemir	1697			
Pratibha Nand, Sushma Drabu and Rajinder K Gupta	1701			
Chemical and Biological Diversity in Fourteen Selections of Four Ocimum Species Bhaskaruni R. Rajeswara Rao, Sushil K. Kothari, Dharmendra K. Rajput, Rajendra P. Patel and Mahendra P. Darokar	1705			
Environmental Effect on Essential Oil Composition of <i>Aloysia citriodora</i> from Corrientes (Argentina) Gabriela Ricciardi, Ana Maria Torres, Ana Laura Bubenik, Armando Ricciardi, Daniel Lorenzo and Eduardo Dellacassa	1711			
Essential Oil of Three Uvaria species from Ivory Coast Koffi A. Muriel, Tonzibo Z. Félix, Gilles Figueredo, Pierre Chalard and Yao T. N'guessan	1715			
Composition of the Essential Oil of <i>Origanum tyttanthum</i> from Tajikistan Farukh S. Sharopov, Muhamadsho A. Kukaniev and William N. Setzer	1719			
Volatiles of French Ferns and "fougère" Scent in Perfumery Didier Froissard, Françoise Fons, Jean-Marie Bessière, Bruno Buatois and Sylvie Rapior	1723			
Volatile Constituents of Two Species of <i>Protium</i> from the Atlantic Rainforest in the State of Pernambuco, Brazil José Gildo Rufino de Freitas, Claudio Augusto Gomes da Camara, Marcílio Martins de Moraes and Henrique Costa Hermenegildo da Silva	1727			
Volatile Constituents of Two Hypericum Species from Tunisia Karim Hosni, Kamel Msaâda, Mouna Ben taârit, Thouraya Chabed and Brahim Marzouk	1727			
Chemical Composition and Possible <i>in vitro</i> Antigermination Activity of Three <i>Hypericum</i> Essential Oils	1751			
Aurelio Marandino, Laura De Martino, Emilia Mancini, Luigi Milella and Vincenzo De Feo	1735			
Antioxidant, Antimicrobial Activities and Fatty Acid Components of Flower, Leaf, Stem and Seed of Hypericum scabrum	1520			
All Sharaghat Composition of Three Essential Oils, and their Mammalian Cell Toxicity and Antimycobacterial Activity	1739			
against Drug Resistant-Tuberculosis and Nontuberculous Mycobacteria Strains Juan Bueno, Patricia Escobar, Jairo René Martínez, Sandra Milena Leal and Elena E. Stashenko	1743			
Antimicrobial and Antioxidant Activities of the Flower Essential Oil of <i>Halimodendron halodendron</i> Jihua Wang, Hao Liu, Haifeng Gao, Jianglin Zhao, Ligang Zhou, Jianguo Han, Zhu Yu and Fuyu Yang	1749			
Composition and Antimicrobial Activity of the Leaf and Twig Oils of <i>Litsea acutivena</i> from Taiwan Chen-Lung Ho. Pei-Chun Liao. Eugene I-Chen Wang and Yu-Chang Su	1755			
Chemical Composition and Antimicrobial Activity of the Volatile Oil from <i>Fusarium tricinctum</i> , the Endophytic				
Ying Zhang, Jianglin Zhao, Jihua Wang, Tijiang Shan, Yan Mou, Ligang Zhou and Jingguo Wang	1759			
Antifungal Activity of Essential Oil from Asteriscus graveolens against Postharvest Phytopathogenic Fungi in Apples Mohamed Znini, Gregory Cristofari, Lhou Majidi, Hamid Mazouz, Pierre Tomi, Julien Paolini and Jean Costa	1763			
Interspecies Comparison of Chemical Composition and Anxiolytic-like Effects of Lavender Oils upon Inhalation Mizuho Takahashi, Tadaaki Satou, Mai Ohashi, Shinichiro Hayashi, Kiyomi Sadamoto and Kazuo Koike	1769			
Essential Oils from the <i>Hyptis</i> genus- A Review (1909-2009) Megil McNeil, Petrea Facey and Roy Porter	1775			

Natural Product Communications 2011

Volume 6, Number 11

Contents

<u>Original Paper</u>	Page
Antifungal Activity of Plumericin and Isoplumericin Dharmendra Singh, Umakant Sharma, Parveen Kumar, Yogesh K. Gupta, M. P. Dobhal and Sarman Singh	1567
A New Diacylated Labdane Diterpenoid from Andrographis wightiana Jalli Madhu Sudhana, Rachakunta Munikishore, Mopuru Vijayabhaskar Reddy, Duvvuru Gunasekar, Alain Blond and Bernard Bodo	1569
Triterpenoid Acids and Lactones from the Leaves of <i>Fadogia tetraquetra</i> var. <i>tetraquetra</i> (Rubiaceae) Dulcie A. Mulholland, Abdelhafeez M.A. Mohammed, Philip H. Coombes, Shafiul Haque, Leena L. Pohjala, Päivi S.M. Tammela and Neil R. Crouch	1573
Isolation of Friedelin from Black Condensate of Cork Ricardo A. Pires, Ivo Aroso, Susana P. Silva, João F. Mano and Rui L. Reis BIO-	1577
Novel Microbial Transformation of Resibufogenin by Absidia coerules Jian Zheng, Dong-hai Su, Dong-sheng Zhang, Xiu-Lan Xin, Jun-ying Liu, Yan Tian, Qing Wei and Xun Cui	1581
Antidiabetic Activity of <i>Terminalia sericea</i> Constituents Nolitha Nkobole, Peter James Houghton, Ahmed Hussein and Namrita Lall	1585
X-ray Crystallographic Study of Ranaconitine Yang Li, Jun-hui Zhou, Gui-jun Han, Min-juan Wang, Wen-ji Sun and Ye Zhao	1589
Obscurine: a New Cyclostachine Acid Derivative from <i>Beilschmiedia obscura</i> Bruno Ndjakou Lenta, Jean Rodolphe Chouna, Pepin Alango Nkeng-Efouet, Samuel Fon Kimbu, Etienne Tsamo and Norbert Sewald	1591
Alkaloids from Papaver coreanum BIODIVERS Dong-Ung Lee, Jong Hee Park, Ludger Wessjohann and Jürgen Schmidt BIODIVERS	1593
Isolation, Structure Elucidation, and Biological Activity of a New Alkaloid from Zanthoxylum rhetsa Karsten Krohn, Stephan Cludius-Brandt, Barbara Schulz, Mambatta Sreelekha and Pottachola Mohamed Shafi	1595
Amplexicine, an Antioxidant Flavan-3-ol from <i>Polygonum amplexicaule</i> Mudasir A. Tantry and Aziz A. Rahman	1597
A New Flavonoid Glycoside from Vaccaria hispanica Haijiang Zhang, Kuiwu Wang, Jie Wu, Yao Chen and Peipei He	1599
Flavonoids from Algerian Endemic <i>Centaurea microcarpa</i> and their Chemotaxonomical Significance Souheila Louaar, Amel Achouri, Mostefa Lefahal, Hocine Laouer, Kamel Medjroubi, Helmut Duddeck and Salah Akkal	1603
On-line (HPLC-NMR) and Off-line Phytochemical Profiling of the Australian Plant, <i>Lasiopetalum macrophyllum</i> Michael Timmers and Sylvia Urban	S1605
Chemical Fingerprint Analysis of Phenolics of <i>Albiziachinensis</i> Based on Ultra-Performance LC-Electrospray Ionization-Quadrupole Time-of-Flight Mass Spectrometry and Antioxidant Activity Abha Chaudhary, Pushpinder Kaur, Neeraj Kumar, Bikram Singh, Shiv Awasthi and Brij Lal	1617
Application to Classification of Mulberry Leaves using Multivariate Analysis of Proton NMR Metabolomic Data Eriko Fukuda, Motoyuki Yoshida, Masaki Baba, Yoshihiro Uesawa, Ryuichiro Suzuki, Osamu Kamo, Koji Tsubono, Kazunori Arifuku, Kazuhisa Yatsunami and Yoshihito Okada	1621
Phenolic Constituents of <i>Knautia arvensis</i> Aerial Parts Jaroslaw Moldoch, Barbara Szajwaj, Milena Masullo, Lukasz Pecio, Wieslaw Oleszek, Sonia Piacente and Anna Stochmal	1627
New Acylated Anthocyanins and Other Flavonoids from the Red Flowers of <i>Clematis</i> Cultivars Masanori Hashimoto, Toshisada Suzuki and Tsukasa Iwashina	1631
Prenylated Isoflavonoids from <i>Rhynchosia edulis</i> Ifedayo V. Ogungbe, Gabrielle M. Hill, Rebecca A. Crouch, Bernhard Vogler, Meenakshi Nagarkoti, William A. Haber and William N. Setzer	1637
Antiparasitic and Antimicrobial Isoflavanquinones from <i>Abrus schimperi</i> Aziz A. Rahman, Volodymyr Samoylenko, Surendra K. Jain, Babu L. Tekwani, Shabana I. Khan, Melissa R. Jacob, Jacob O. Midiwo, John P. Hester, Larry A. Walker and Ilias Muhammad	1645
Two New Rotenoids from <i>Boerhavia repens</i> Mamona Nazir, Muhammad Saleem, Naheed Riaz, Maria Hafeez, Misbah Sultan, Abdul Jabbar and Muhammad Shaiq Ali	1651
A Comparison of the Diastereoisomers, Silybin A and Silybin B, on the Induction of Apoptosis in K562 cells Jiyong Zhang, Qiuying Luan, Yanze Liu, David Y-W Lee and Zhao Wang	1653

(Continued inside back cover)